

ORIGINAL ARTICLE

Gondwana breakup under the ephemeral look

Felipe Donateli Gatti¹  | Frederico Falcão Salles^{1,2}  | Phillip John Suter³  |
Yuri Luiz Reis Leite¹ ¹Programa de Pós-Graduação em Ciências
Biológicas, Universidade Federal do
Espírito Santo, Vitória, Brazil²Museu de Entomologia, Universidade
Federal de Viçosa, Viçosa, Brazil³Department of Ecology, Environment
and Evolution, La Trobe University,
Melbourne, Vic., Australia

Correspondence

Felipe D. Gatti, Universidade Federal de
Espírito Santo, Vitória, Brazil.
Email: gattifd@gmail.com

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Abstract

Atalophlebiinae (Ephemeroptera, Leptophlebiidae) is a mayfly subfamily present in temperate and mountainous areas of South America and Australia. We tested the hypothesis that both vicariance and dispersal related to the second phase of Gondwana breakup—which began in the Early Cretaceous and resulted in the separation between Madagascar and India from Antarctica and Australia—contributed to the origin, diversification, and shaped the current distribution of this group. The hypothesis was tested using Bayesian phylogenetic trees, fossil-based molecular dating, and ancestral range estimation to reconstruct the biogeography of the lineages within this group. The results suggested an origin in the late Gondwana supercontinent for Atalophlebiinae (85.76–136.63 mya) after a vicariant event during the Cretaceous period. Subsequently, the lineage diversified into a scenario that refers to a Gondwanic corridor formed by South America, Antarctica, and Australia. At the end of the separation of the continents that made up the Gondwanic corridor, speciation occurred within the current distribution areas. The diversity and current distribution of Atalophlebiinae were shaped by complex processes of vicariance, dispersal, and speciation within the Gondwanic corridor during the second phase of the supercontinent breakup. Mayflies have difficulty in crossing transoceanic barriers, which suggests that most living taxa are the result of more recent local ecological and historical processes.

KEYWORDS

ancestral area, dispersal, divergence times, historical biogeography, vicariance

1 | INTRODUCTION

Vicariance and dispersal associated with plate tectonics helped shape diversification and distribution patterns of biodiversity across the planet (Chamberland et al., 2018; Jurado-Rivera et al., 2017; Toussaint, Hendrich, et al., 2017). Gradual breakup of ancient supercontinents contributed to cladogenetic events, dividing lineages by vicariance (see Kim & Farrell, 2015; Sanmartín & Ronquist, 2004; Toussaint et al., 2017) and dispersal through corridors that connected land fragments (see Reguero et al., 2014; Seton et al., 2012). Thus, vicariant events associated with the Gondwana breakup are commonly invoked to explain the disjunct distribution of ancient lineages across continents in the southern hemisphere (McCulloch

et al., 2016; Sanmartín & Ronquist, 2004), while transoceanic dispersal would be responsible for this pattern of distribution in more recent lineages (Condamine et al., 2013; Martín-Bravo & Daniel, 2016).

Gondwana breakup occurred gradually and can be divided into two phases. The first began in the Early Jurassic (~180 mya), resulting in the separation of West Gondwana (South America/Africa) from East Gondwana (Madagascar, India, Antarctica, and Australia), ca. 140 mya (Mueller & Jokat, 2019; Seton et al., 2012; Thompson et al., 2019). However, it is important to note that southern South America was connected to the Antarctic Peninsula through the Weddellian Isthmus until the opening of Drake Passage at ca. 35 mya (Elsworth et al., 2017). The second phase began in the Early Cretaceous (~135 mya), and resulted in the separation between Madagascar/

India from Antarctica/Australia forming a corridor that connected fragments from South America, Antarctica, and Australia at ca. 100 mya (Gibbons et al., 2013; Seton et al., 2012; Seton et al., 2012; Thompson et al., 2019). Concurrently to these events, rupture between Africa and South America and separation of Madagascar from India took place (Gibbons et al., 2013; Seton et al., 2012; Thompson et al., 2019).

Atalophlebiinae (Ephemeroptera, Leptophlebiidae) are a cool-adapted mayfly subfamily with amphinotic distribution, present in temperate and mountainous areas of South America and Australia (Monjardim et al., 2020; O'Donnell & Jockusch, 2008; Pescador & Peters, 1980; Savage, 1987). Previously, researchers also considered taxa from tropical areas of South America, Africa, and Madagascar as part of this group, suggesting an ancient Gondwana origin of this subfamily (Kluge, 2009; Pescador & Peters, 1980; Savage, 1987). However, recent studies indicate a monophyletic lineage of only amphinotic taxa (Monjardim et al., 2020; O'Donnell & Jockusch, 2008). Ephemeroptera is one of the oldest insect lineages (Misof et al., 2014), and its amphinotic distribution pattern is recurrent in other taxa of the group, such as Ameletopsidae, Coloburiscidae, Nesameletidae, and Oniscigastridae (see Edmunds, 1972; Sartori & Brittain, 2015). Considering the role of plate tectonics in biogeography and our current knowledge of the cool-adapted mayflies, here, we tested the hypothesis that Atalophlebiinae originated and diversified under the influence of both vicariant and dispersal events, during the second phase of Gondwana breakup, which would explain the disjunct distribution between South American and Australian taxa and their absence in tropical Africa, Madagascar, and Indo-Malayan regions.

2 | MATERIALS AND METHODS

2.1 | Taxon sampling and molecular dataset

We sequenced 76 specimens (Table S1) for two molecular markers (Table S2): the D2–D5 region of the 28S ribosomal RNA gene (Gillespie et al., 2004, 2005) and a partial region of the *Cytochrome c oxidase subunit 1* gene (Folmer et al., 1994). We also used 59 sequences available on GenBank to complement our dataset (Table S1). Nine outgroup genera were selected based on a recent phylogeny of the Ephemeroptera (Monjardim et al., 2020; Ogden et al., 2019; O'Donnell & Jockusch, 2008) and the availability of fossils for calibration. Our data matrix comprised 19 genera, representing approximately 55% of the generic composition of Atalophlebiinae. All specimens sequenced in this research are stored in the Museu de Entomologia of the Universidade Federal de Viçosa, under the care of the authors (FFS) or in the Coleção Zoológica Norte Capixaba, Universidade Federal do Espírito Santo, Brazil.

2.2 | Phylogenetic analyses

Sequences were aligned in Geneious 9.0 (www.geneious.com), and nucleotide substitution models for each marker were selected using

the Corrected Akaike Information Criterion (AICc) in jModelTest2 (Darriba et al., 2012) on CIPRES (Miller et al., 2010). Saturation level of sequences was verified by Xia's test (Xia et al., 2003) in DAMBE 7 (Xia, 2018), and the third codon position of COI was consequently excluded from the analyses. The concatenated molecular data matrix comprised 1622 base pairs (1196 bp from 28S and 426 bp from COI) for 97 operational taxonomic units (Alignment S1). Models selected for each partition were GTR+G to 28S and TrN+G to COI (Table S3).

Phylogenetic tree was inferred using Bayesian inference in MrBayes 3.2.7a (Ronquist et al., 2012) on CIPRES (Miller et al., 2010). Eight Markov chain Monte Carlo (MCMC) iterations were run simultaneously for 1.58 million generations with sampling trees every 1000 generations and 25% of burn-in, until the convergence diagnostic reached the stop value (standard deviation of split frequencies <0.01). Support of nodes was provided by posterior probabilities (PP) as directly estimated from the majority rule consensus topology. Considering recent discussions about statistical significance (Amrhein et al., 2019; Hurlbert et al., 2019; Pike, 2019; Wasserstein et al., 2019), the logic, background knowledge, and experimental design were considered alongside PP to reach a conclusion and decide on its certainty. Therefore, nodes with PP value higher than 0.85 were considered well-supported.

2.3 | Divergence times

We used the relaxed uncorrelated lognormal molecular clock with a tree prior using the birth-death incomplete sampling algorithm (Stadler, 2009). Substitution models for each partition (28S and COI) were selected according to AICc (Table S3), and monophyly was forced based on BI results. Clock points calibration was based on seven date priors based on fossils ($n = 5$), geological event and probabilities of dispersal ($n = 1$, adapted from Landis, 2017), and secondary data derived from previous analyses ($n = 1$).

(1) Root was calibrated to represent the minimum and maximum (242–290 mya) ages of fossil species *Protereisma permianum* Sellards 1907 (Protereismatidae), believed to be one of the stem groups of Ephemeroptera (Godunko et al., 2011; Grimaldi & Engel, 2005; Sroka et al., 2015), and the mayfly *Triassonurus doliiformis* Sinitshenkova & Papier, 2005 (Siphonuridae), the lineage with the oldest origin in our dataset (Lognormal distribution, offset = 242.0, Mean = 8.5, Standard deviation = 1.0, mean in real space). (2) Oligoneuriidae initial diversification was calibrated based on *Incogemina nubile* Storari et al., 2020. (Oligoneuriidae) (Lognormal, offset = 112.6, $M = 30.0$, $S = 1.0$, mean in real space). (3) Leptophlebiidae initial diversification was calibrated based on the age of ~175 mya given by Grimaldi and Engel (2005) for origin of the family (Normal distribution, Mean=175.0, Sigma=25.0). (4) Leptophlebiinae initial diversification was calibrated based on the fossil *Aureophlebia sinitshenkova* Peters & Peters 2000 (Leptophlebiinae) (Lognormal, offset = 89.3, $M = 20.0$, $S = 1.0$, mean in real space). (5) *Paraleptophlebia* initial diversification was calibrated based on the fossil *Paraleptophlebia prisca* (Pictet & Hagen, 1856) (Lognormal, offset = 33.9, $M = 13.0$, $S = 1.0$, mean in real space). (6) Calibration of the most recent common ancestor of Atalophlebiinae

from the lineage that originated *Radima* Akers, Peters & Peters, 2003 (Lognormal, offset = 85.0, $M = 21.0$, $S = 1.0$, mean in real space) considered the final period of separation between Madagascar/India and Antarctica/Australia (~100 mya, Gibbons et al., 2013; Seton et al., 2012; Thompson et al., 2019; White et al., 2013) and low probability of dispersal between these areas after separation (see supplementary data of Landis, 2017; Sanmartín & Ronquist, 2004). 7) Divergence between *Atalophlebia* Eaton, 1881, and *Atalomicria* Harker, 1954, was calibrated with the fossil *Atalophlebia culleni* Etheridge & Olliff 1890 (Lognormal, offset = 2.6, $M = 20.0$, $S = 1.0$, mean in real space). Fossil information can be accessed in the Fossilworks Paleobiology Database (<http://fossilworks.org>).

The input file was constructed in BEAUti v2.5.2 and run in BEAST2 v2.5.2 (Bouckaert et al., 2019) for 100 million generations and trees sampled every 10,000 generations. The convergences of runs and the effective sample size ($ESS > 1000$) of parameters were examined in Tracer v1.7.1 (Rambaut et al., 2018). A tree with maximum clade credibility topology, using a burn-in of 25%, was constructed with TreeAnnotator v2.5.2, and analyses were run on CIPRES (Miller et al., 2010).

2.4 | Ancestral area reconstruction

As suggested by Ree and Sanmartín (2018), the model for reconstruction of ancestral areas was chosen to consider the structure and assumptions of models, and not a statistical method that assumes probabilistic equivalence between different models (e.g., AIC). We selected Dispersal-Extinction-Cladogenesis (DEC; Ree et al., 2005), which is a model that allows the incorporation of fossil and geological information, and to co-estimate phylogeny as a stochastic process in continuous time and incorporates both vicariance and dispersal (Ree & Sanmartín, 2018; Ree & Smith, 2008; Ronquist & Sanmartín, 2011).

Genera occurrences were defined by presence or absence in South America (A), Madagascar (B), Australia (C), and New Zealand (D). Gondwana breakup is well documented (see Bache et al., 2014; Elsworth et al., 2017; Mueller & Jokat, 2019; Seton et al., 2012; White et al., 2013), and the probability of dispersal was assigned according to the availability of connections between areas across four time slices: (t1) 108 to 85 mya; (t2) 85 to 50 mya; (t3) 50 to 35 mya; and (t4) 35 to 0 mya. As suggested by Landis (2017), the probability of dispersal (constrained to sum to 1) was attributed to short-distances dispersal ($s = 0.7$); medium-distances dispersal ($m = 0.2$); and long-distances dispersal ($l = 0.1$). Thus, probability of medium is implied to exist in short, and long-distance dispersal is implied to exist between all area pairs. Therefore, short distance has value 1 ($s+m+l$) and medium distance has value 0.3 ($m+l$) (Table S4; adapted from Landis, 2017).

Time slices and dispersal probabilities considered five geological events: the separation between Madagascar/India and Antarctica associated with low probability of dispersal after separation (~85 mya, supplementary data of Landis, 2017; White et al., 2013; Gibbons et al., 2013; Sanmartín & Ronquist, 2004); Tasman

Sea opening (~50 mya, Bache et al., 2014; White et al., 2013); the opening of Drake Passage and Tasman Gateway with consequent Antarctica glaciation (~35 mya, Elsworth et al., 2017; Scher et al., 2015). The analyses were run using the "BioGeoBEARS" package (Matzke, 2014) on R (R Core Team, 2020) under the RASP interface (Yu et al., 2015). The resulting phylogeny from BEAST2 was used as a guide tree (consensus tree).

3 | RESULTS

Bayesian inference recovered Atalophlebiinae as monophyletic with high support (Figures 1 and S1) and revealed new phylogenetic relationships among genera in internal clades. *Garinjuga* Campbell & Suter, 1988, appears as sister to all other Atalophlebiinae genera, including *Massartella* Lestage, 1930, which, in turn, is sister to the remaining Atalophlebiinae. Two other clades contain genera from South America and Australia (Figure 1).

Estimated age for the origin of Atalophlebiinae was 107.83 mya [median age, 95% highest posterior density interval (95% HPD): 85.76–136.63, Figure 1, Table 1, node 1], with initial diversification at 94.52 mya (68.53–123.44, Figure 1, Table 1, node 2). Thus, both the origin and initial lineage divergence occurred in the Cretaceous, during the second phase of Gondwana breakup. The result suggests that most living genera originated within their current distribution area.

Inference of ancestral areas (Figure 2, Table 1) suggested that Atalophlebiinae originated in the Gondwana supercontinent after a vicariant event (node 1). Subsequently, this lineage diversified into a scenario that refers to the Gondwanic corridor formed by South America, Antarctica, and Australia (node 2). Then, vicariance again separated the lineage leading to *Massartella* from the remaining Atalophlebiinae at ca. 85 mya (node 3). Other Australian (*Jappa* Harker, 1954 and the clade *Atalomicria* + *Atalophlebia*) and South American (*Penaphlebia* Peters & Edmunds, 1972 and the clade *Massartellopsis* Demoulin, 1955 + *Meridialis* Peters & Edmunds, 1972) sister lineages resulted from vicariant events at ca. 57 mya (nodes 5, 9).

4 | DISCUSSION

The topology of Atalophlebiinae herein proposed contains four main lineages: *Garinjuga* from Australia, *Massartella* from South America, and two others with genera from South America and Australia region (Figures 1, 2, and S1). *Garinjuga* samples and new sequences addition (Table S1) helped to establish the relationship among main lineages of the group, which in previous research were unclear (see Monjardim et al., 2020). *Jappa* and *Austrophlebioides* Campbell & Suter, 1988, belong to the clade supported by node 8 (Figures 1 and 2) diverging from the results of Monjardim et al., (2020). This study did not recover any previously proposed clades within Atalophlebiinae based on morphological data (see Christidis, 2006;

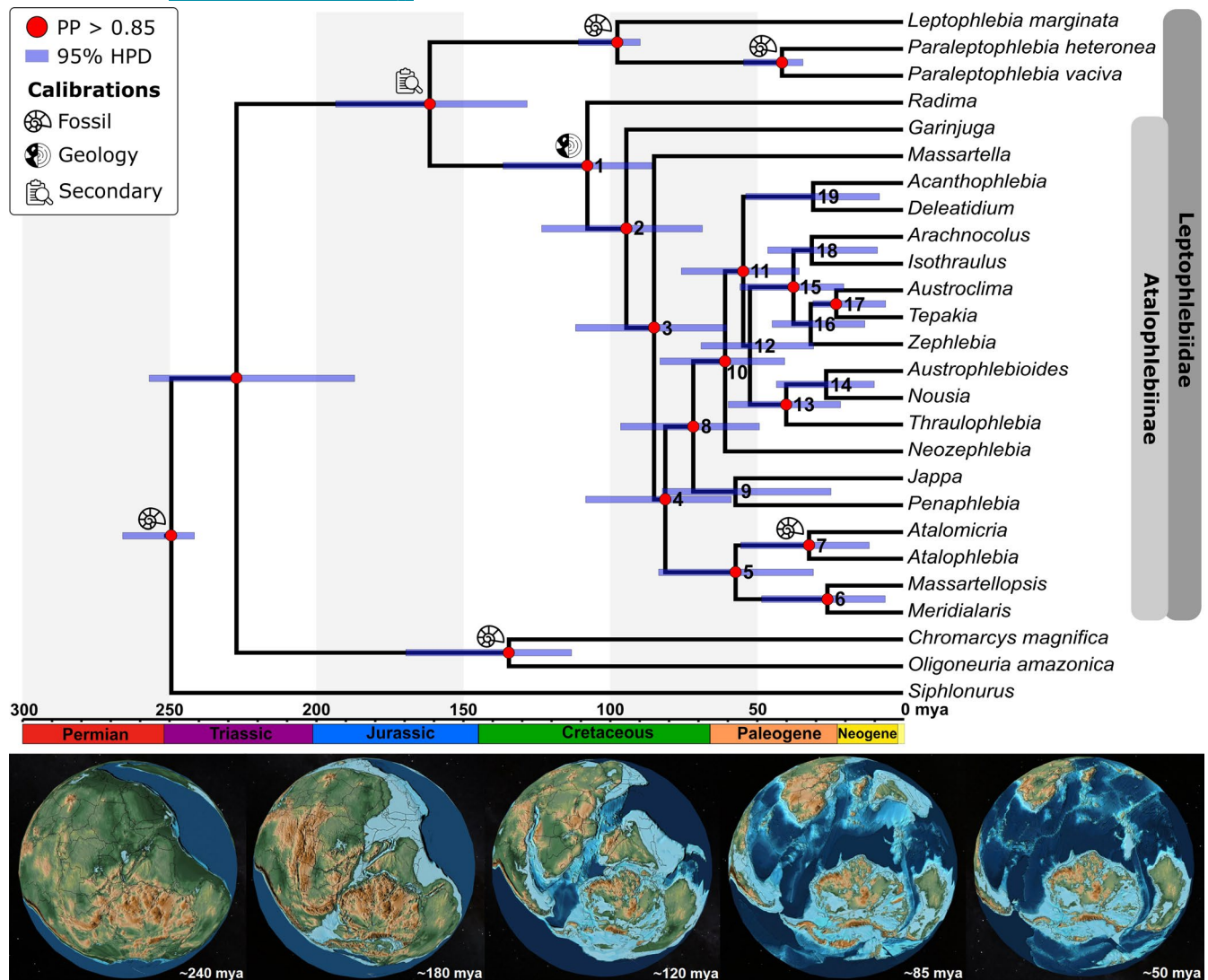


FIGURE 1 Time-calibrated [millions of years ago (mya)] phylogeny for Atalophlebiinae. Posterior probabilities (PP) referring to Bayesian inference tree (Figure S1). HPD: highest posterior density. Nodes used to calibrate tree were identified with specific symbols according to type of calibration (top-left box). Node numbers refer to Figure 2 and Table 1. Palaeogeographical maps reconstructed by Scotese's PALEOMAP Project available in Global Geology (Scotese & Dreher, 2012)

Finlay & Bae, 2008; Pescador & Peters, 1980). Considering that they occur in similar environments throughout their distribution, and therefore experience similar ecological filters, many species may have evolved similar characteristics independently (see Bower & Winemiller, 2019). This indicates that some morphological similarities shared between Atalophlebiinae genera may be the result of evolutionary convergence.

The results suggest that the most recent common ancestor between Atalophlebiinae and the lineage that originated the Malagasy group (here represented by *Radima*) lived in Gondwana in the Cretaceous period (Figures 1 and 2, Table 1, node 1). These lineages diverged allopatrically during the second phase of the supercontinent breakup (Gibbons et al., 2013; Seton et al., 2012; Thompson et al., 2019), and the vicariant event that promoted speciation is probably related to the separation between Madagascar/India from Antarctica/Australia. The breakup process between these areas

began at ca. 135 mya, with the migration of the Indian Plate, culminating in the opening of the Indian Ocean at ca. 100 mya (Gibbons et al., 2013; Seton et al., 2012; Thompson et al., 2019).

The initial divergence in Atalophlebiinae occurs in a scenario that refers to the Gondwanian corridor formed by South America, Antarctica, and Australia (Reguero et al., 2014; Seton et al., 2012; Thompson et al., 2019) during the Upper Cretaceous (Figures 1 and 2, Table 1, node 2). The ancestor of the *Garinjuga* lineage was probably limited to the geographic area where the divergence occurred (area C), which can be explained, for example, by a peripatric speciation, while the remaining lineage (node 3) inherits the entire ancestral range and probably increases its distribution by dispersal within that range (scenario 3 in Ree et al., 2005). Subsequently, a vicariant event isolated an ancestral population (node 3) in South America that gave rise to the *Massartella* lineage. Changes in sea level, associated with temperature increase on

TABLE 1 Combined results of divergence time and ancestral area evolution of Atalophlebiinae based on BEAST2 time tree and Dispersal-Extinction-Cladogenesis (DEC)

Clade	Divergence time (mya)		DEC		
	Median	95% HPD	Ancestral area	Probabilities	Event
Root	249.95	242.09–266.57	—	—	—
Oligoneuriidae + Leptophlebiidae	227.69	187.32–257.53	—	—	—
Oligoneuriidae (Chromarcys + Oligoneuria)	134.62	113.24–169.86	—	—	—
Leptophlebiidae	161.68	128.42–193.84	—	—	—
Leptophlebiinae (<i>Leptophlebia</i> + <i>Paraleptophlebia</i>)	97.58	89.77–110.9	—	—	—
<i>Paraleptophlebia</i>	41.36	34.19–54.5	—	—	—
Node 1	107.83	85.76–136.63	ABC	0.47	Vicariance
Node 2	94.52	68.53–123.44	AC	0.56	Dispersal
Node 3	85.05	60.11–111.87	AC	0.45	Vicariance
Node 4	81.21	58.67–108.4	C	0.39	Dispersal
Node 5	57.18	30.64–83.5	AC	1	Vicariance
Node 6	25.83	6.09–48.3	A	1	s. w. a
Node 7	32.1	11.53–55.45	C	1	s. w. a
Node 8	71.66	49.07–96.49	CD	0.33	Ambiguous
Node 9	57.24	24.60–82.25	AC	1	Vicariance
Node 10	60.79	40.42–83.02	D	0.84	Dispersal
Node 11	54.6	35.45–75.74	CD	0.83	Dispersal
Node 12	52.3	30.55–69.01	CD	0.96	Vicariance
Node 13	39.9	21.37–59.72	C	0.94	Dispersal
Node 14	26.26	9.83–43.26	AC	0.58	Dispersal
Node 15	37.44	20.24–55.71	D	1	s. w. a
Node 16	31.55	13.05–44.72	D	1	s. w. a
Node 17	22.8	5.95–30.78	D	1	s. w. a
Node 18	31.17	8.74–46.21	D	1	s. w. a
Node 19	30.7	8.05–53.74	D	1	s. w. a

Note: Node numbers refer to Figures 1 and 2.

Abbreviations: HPD, highest posterior density; s.w.a., speciation within area.

the planet, culminated in several cycles of marine transgressions in Patagonia during the Upper Cretaceous and Paleocene (Haq, 2014; Le Roux, 2012a; Malumian & Nanez, 2011; Parras & Griffin, 2013), which could have prevented dispersal events between South America and the Antarctic Peninsula. In addition, the temperature and precipitation calculated for these periods indicates a climate warm and humid subtropical temperate in this region (Le Roux, 2012b; Varela et al., 2018). This fact may have induced mayfly populations to seek colder habitats in mountains, leaving the lowlands, which could also have promoted allopatric speciation. Events of the same nature could have also been responsible for isolating the lineages that originated *Atalophlebia* + *Atalomicria* and *Jappa* in Australia from *Massartellopsis* + *Meridialaris* and *Penaphlebia* in South America (node 5 and 9). Thus, marine transgressions and/or climate changes were probably strong enough to isolate Atalophlebiinae populations, as this group is adapted to

cold streams and rivers and is extremely intolerant of saltwater (Dos Santos et al., 2018).

Massartella descended from the oldest split which produced one lineage presently restricted to South America. Nowadays, it has a wide and disjunct distribution on the tabletops of the Pantepui region and in the mountains of the Atlantic Forest (Dominguez et al., 2006; Pescador & Peters, 1990). *Massartellopsis* and *Meridialaris* speciated during the uplift of the Andes (Figures 1 and 2, Table 1, node 6), a region where they are currently found, but where *Massartella* does not occur (Derka et al., 2009; Dominguez et al., 2006; Hoorn et al., 2010).

New Zealand emerges as an important factor in the diversification of Atalophlebiinae, after events of unclear nature (Figures 1 and 2, Table 1, node 4 and 8), possibly related to extinction events (or with sample gap), during the initial period of its separation from Australia (~85 mya) until the opening of the Tasman Sea (~50 mya, see Bache

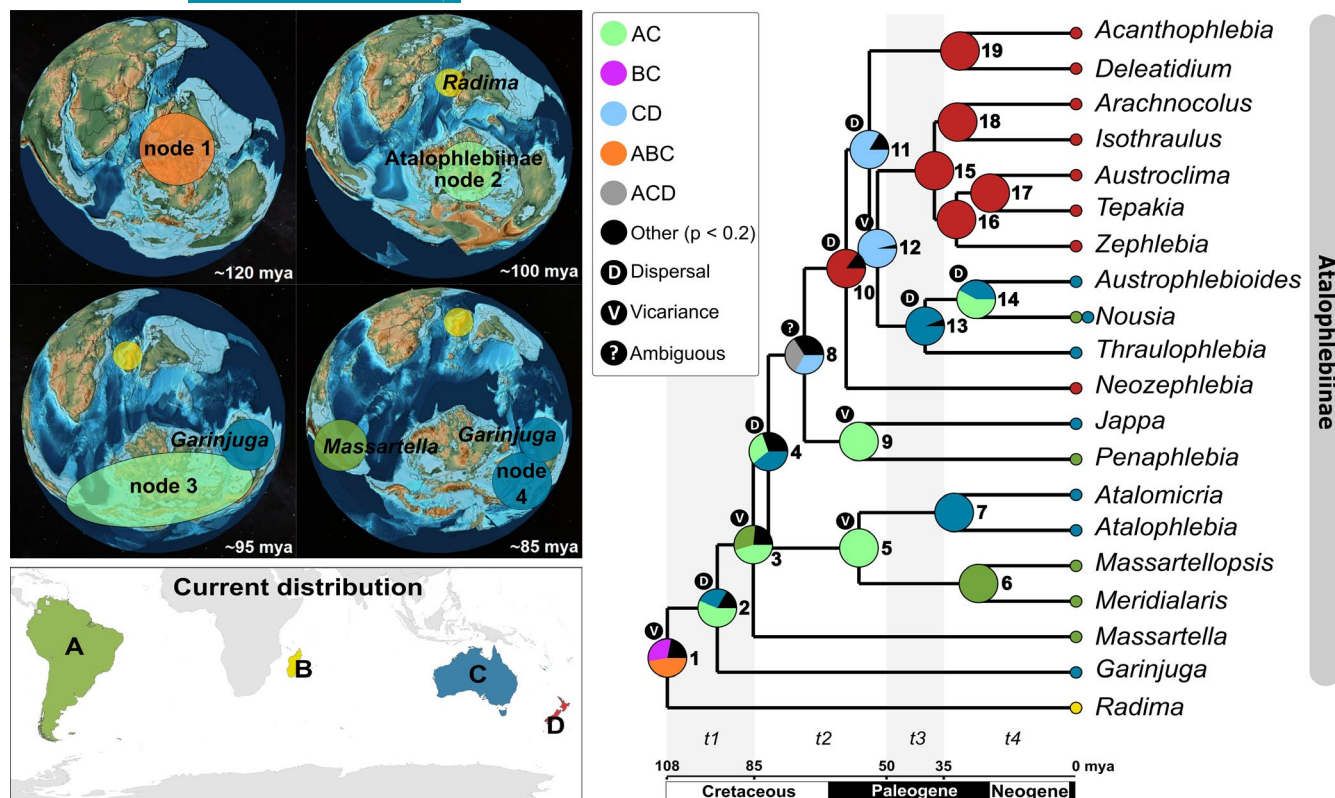


FIGURE 2 Reconstruction of ancestral area for Atalophlebiinae using Dispersal-Extinction-Cladogenesis (DEC) model. Palaeogeographical maps show probable scenarios of origin and initial diversification of the group. Nodes were colored according to the areas with highest relative probability inferred by DEC. Node numbers refer to Figure 1 and Table 1. The lower-left map represents the three areas (A, C, D) and Madagascar (for *Radima* outgroup) of current genera distribution implemented combined with time slices (t_1 , t_2 , t_3 , and t_4) in the matrix of dispersal probabilities (Table S4). Palaeogeographical maps reconstructed by Scotese's PALEOMAP Project available in Global Geology (Scotese & Dreher, 2012)

et al., 2014). An ancestral population isolated in this region (node 10) was the origin of the lineage of *Neozephlebia* Penniket, 1961, while another lineage increased its range by dispersal to Australia (node 11). Subsequently, peripheral speciation formed the lineage (node 19) in New Zealand region, while others (node 12) inherited the entire ancestral range and increases distribution by dispersal (scenario 3 in Ree et al., 2005). Later, that lineage (node 12) was divided by a vicariant event, probably related to landscape changes during the opening of the Tasman Sea and Tasmanian Gateway at ca. 50–35 mya (see Bache et al., 2014; Scher et al., 2015). After the separation of the Gondwanic corridor (~35 mya, Elsworth et al., 2017; Scher et al., 2015; Seton et al., 2012), speciation occurred within the current distribution areas, indicating that Atalophlebiinae was unable to disperse across transoceanic barriers and suggests that most living taxa are the result of more recent local historical and ecological processes.

The biogeographic history of Atalophlebiinae is congruent with events that occurred during the second phase of the gradual process of Gondwana breakup in the Cretaceous and Paleogene. Our results provided evidence that vicariance and dispersal both played roles in the history of diversification prior to the completion of the second

phase of Gondwana breakup. This pattern, together with natural extinction processes and its low dispersal capacity across transoceanic barriers, may explain its absence in other regions that form the circum-Antarctic pattern, such as tropical Africa, Madagascar, and India.

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CONFLICT OF INTEREST

The authors declare no conflicts of interest.

ORCID

Felipe Donatelli Gatti  <https://orcid.org/0000-0003-0926-4670>

Frederico Falcão Salles  <https://orcid.org/0000-0001-8331-5929>

Phillip John Suter  <https://orcid.org/0000-0002-6116-2885>

Yuri Luiz Reis Leite  <https://orcid.org/0000-0002-0248-8738>

REFERENCES

- Amrhein, V., Greenland, S., & McShane, B. (2019). Scientists rise up against statistical significance. *Nature*, 567, 305–307. <https://doi.org/10.1038/d41586-019-00857-9>.
- Bache, F., Mortimer, N., Sutherland, R., Collot, J., Rouillard, P., Stagpoole, V., & Nicol, A. (2014). Seismic stratigraphic record of transition from Mesozoic subduction to continental breakup in the Zealandia sector of eastern Gondwana. *Gondwana Research*, 26, 1060–1078. <https://doi.org/10.1016/j.gr.2013.08.012>.
- Bouckaert, R., Vaughan, T. G., Barido-Sottani, J., Duchêne, S., Fourment, M., Gavryushkina, A., Heled, J., Jones, G., Kühnert, D., De Maio, N., Matschiner, M., Mendes, F. K., Müller, N. F., Ogilvie, H. A., du Plessis, L., Poppinga, A., Rambaut, A., Rasmussen, D., Siveroni, I., ... Drummond, A. J. (2019). BEAST 2.5: An advanced software platform for Bayesian evolutionary analysis. *PLoS Computational Biology*, 15, e1006650. <https://doi.org/10.1371/journal.pcbi.1006650>.
- Bower, L. M., & Winemiller, K. O. (2019). Fish assemblage convergence along stream environmental gradients: An intercontinental analysis. *Ecography*, 42, 1691–1702. <https://doi.org/10.1111/ecog.04690>.
- Chamberland, L., McHugh, A., Kechejian, S., Binford, G. J., Bond, J. E., Coddington, J., Dolman, G., Hamilton, C. A., Harvey, M. S., Kuntner, M., & Agnarsson, I. (2018). From Gondwana to GAAR landia: Evolutionary history and biogeography of ogre-faced spiders (*Deinopis*). *Journal of Biogeography*, 45, 2442–2457. <https://doi.org/10.1111/jbi.13431>.
- Christidis, F. (2006). Phylogenetic relationships of the Australian Leptophlebiidae (Ephemeroptera). *Invertebrate Systematics*, 19, 531–539. <https://doi.org/10.1071/IS05022>.
- Condamine, F. L., Sperling, F. A., & Kergoat, G. J. (2013). Global biogeographical pattern of swallowtail diversification demonstrates alternative colonization routes in the Northern and Southern hemispheres. *Journal of Biogeography*, 40, 9–23. <https://doi.org/10.1111/j.1365-2699.2012.02787.x>.
- Darriba, D., Taboada, G. L., Doallo, R., & Posada, D. (2012). jModelTest 2: More models, new heuristics and parallel computing. *Nature Methods*, 9, 772. <https://doi.org/10.1038/nmeth.2109>.
- Derka, T., Svitok, M., & Schlögl, J. (2009). *Massartella hirsuta* sp. nov. (Ephemeroptera: Leptophlebiidae: Atalophlebiinae) and new data on mayflies of Guyana Highlands. *Aquatic Insects*, 31, 83–94. <https://doi.org/10.1080/01650420902811992>.
- Domínguez, E., Molineri, C., Pescador, M. L., Hubbard, M. D., & Nieto, C. (2006). *Ephemeroptera de América del Sur* (Vol. 2). Pensoft Publishers.
- Dos Santos, D. A., Molineri, C., Nieto, C., Zuñiga, M. C., Emmerich, D., Fierro, P., Pessacq, P., Rios-Touma, B., Márquez, J., Gomez, D., Salles, F. F., Encalada, A. C., Príncipe, R., Gómez, G. C., Valdovinos Zarges, C., & Domínguez, E. (2018). Cold/Warm stenothermic freshwater macroinvertebrates along altitudinal and latitudinal gradients in Western South America: A modern approach to an old hypothesis with updated data. *Journal of Biogeography*, 45, 1571–1581. <https://doi.org/10.1111/jbi.13234>.
- Edmunds, G. F. (1972). Biogeography and evolution of Ephemeroptera. *Annual Review of Entomology*, 17, 21–42. <https://doi.org/10.1146/annurev.en.17.010172.000321>.
- Elsworth, G., Galbraith, E., Halverson, G., & Yang, S. (2017). Enhanced weathering and CO₂ drawdown caused by latest Eocene strengthening of the Atlantic meridional overturning circulation. *Nature Geoscience*, 10, 213–216. <https://doi.org/10.1038/ngeo2888>.
- Finlay, K. J., & Bae, Y. J. (2008). Phylogenetic relationships of the Australian Leptophlebiidae. In F. R. Hauer, J. A. Stanford, & R. L. Newell (Eds.), *International advances in the ecology, zoogeography and systematics of mayflies and stoneflies* (pp. 233–275). University of California Press.
- Folmer, O., Black, M., Hoeh, W., Lutz, R., & Vrijenhoek, R. (1994). DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology & Biotechnology*, 3, 294–297.
- Gibbons, A. D., Whittaker, J. M., & Müller, R. D. (2013). The breakup of East Gondwana: Assimilating constraints from Cretaceous ocean basins around India into a best-fit tectonic model. *Journal of Geophysical Research: Solid Earth*, 118, 808–822. <https://doi.org/10.1002/jgrb.50079>.
- Gillespie, J., Cannone, J., Gutell, R., & Cognato, A. (2004). A secondary structural model of the 28S rRNA expansion segments D2 and D3 from rootworms and related leaf beetles (Coleoptera: Chrysomelidae: Galerucinae). *Insect Molecular Biology*, 13, 495–518. <https://doi.org/10.1111/j.0962-1075.2004.00509.x>.
- Gillespie, J. J., Munro, J. B., Heraty, J. M., Yoder, M. J., Owen, A. K., & Carmichael, A. E. (2005). A secondary structural model of the 28S rRNA expansion segments D2 and D3 for chalcidoid wasps (Hymenoptera: Chalcidoidea). *Molecular Biology and Evolution*, 22, 1593–1608. <https://doi.org/10.1093/molbev/msi152>.
- Godunko, R., Staniczek, A., & Bechly, G. (2011). Coxopteroptera, a new fossil order of Palaeoptera (Arthropoda: Insecta), with comments on the phylogeny of the stem group of mayflies (Ephemeroptera). *Insect Systematics & Evolution*, 42, 101–138. <https://doi.org/10.1163/187631211X578406>.
- Grimaldi, D., & Engel, M. S. (2005). *Evolution of the Insects*. Cambridge University Press.
- Hag, B. U. (2014). Cretaceous eustasy revisited. *Global and Planetary Change*, 113, 44–58. <https://doi.org/10.1016/j.gloplacha.2013.12.007>.
- Hoorn, C., Wesselingh, F. P., ter Steege, H., Bermudez, M. A., Mora, A., Sevink, J., Sanmartin, I., Sanchez-Meseguer, A., Anderson, C. L., Figueiredo, J. P., Jaramillo, C., Riff, D., Negri, F. R., Hooghiemstra, H., Lundberg, J., Stadler, T., Sarkinen, T., & Antonelli, A. (2010). Amazonia through time: Andean uplift, climate change, landscape evolution, and biodiversity. *Science*, 330, 927–931. <https://doi.org/10.1126/science.1194585>.
- Hurlbert, S. H., Levine, R. A., & Utts, J. (2019). Coup de grâce for a tough old bull: “Statistically significant” expires. *The American Statistician*, 73, 352–357.
- Jurado-Rivera, J. A., Pons, J., Alvarez, F., Botello, A., Humphreys, W. F., Page, T. J., Iliffe, T. M., Willassen, E., Meland, K., Juan, C., & Jaime, D. (2017). Phylogenetic evidence that both ancient vicariance and dispersal have contributed to the biogeographic patterns of anchialine cave shrimps. *Scientific Reports*, 7, 1–11. <https://doi.org/10.1038/s41598-017-03107-y>.
- Kim, S. I., & Farrell, B. D. (2015). Phylogeny of world stag beetles (Coleoptera: Lucanidae) reveals a Gondwanan origin of Darwin's stag beetle. *Molecular Phylogenetics and Evolution*, 86, 35–48. <https://doi.org/10.1016/j.ympev.2015.02.015>.
- Kluge, N. J. (2009). Higher system of Atalophlebiinae (Leptophlebiidae) with description of three new species of Terpidia sl from Peruvian Amazonia. *Russian Entomological Journal*, 18, 243–256.
- Landis, M. J. (2017). Biogeographic dating of speciation times using paleogeographically informed processes. *Systematic Biology*, 66, 128–144. <https://doi.org/10.1093/sysbio/syw040>.
- Le Roux, J. P. (2012a). A review of Tertiary climate changes in southern South America and the Antarctic Peninsula. Part 1: Oceanic conditions. *Sedimentary Geology*, 247, 1–20. <https://doi.org/10.1016/j.sedgeo.2011.12.014>.
- Le Roux, J. P. (2012b). A review of Tertiary climate changes in southern South America and the Antarctic Peninsula. Part 2: Continental

- conditions. *Sedimentary Geology*, 247, 21–38. <https://doi.org/10.1016/j.sedgeo.2011.12.001>.
- Malumian, N., & Nanez, C. (2011). The Late Cretaceous-Cenozoic transgressions in Patagonia and the Fuegian Andes: Foraminifera, palaeoecology, and palaeogeography. *Biological Journal of the Linnean Society*, 103, 269–288. <https://doi.org/10.1111/j.1095-8312.2011.01649.x>.
- Martín-Bravo, S., & Daniel, T. F. (2016). Molecular evidence supports ancient long-distance dispersal for the amphi-Atlantic disjunction in the giant yellow shrimp plant (*Barleria oenotheroides*). *American Journal of Botany*, 103, 1103–1116. <https://doi.org/10.3732/ajb.1600083>.
- Matzke, N. J. (2014). Model selection in historical biogeography reveals that founder-event speciation is a crucial process in island clades. *Systematic Biology*, 63, 951–970. <https://doi.org/10.1093/sysbio/syu056>.
- McCulloch, G. A., Wallis, G. P., & Waters, J. M. (2016). A time-calibrated phylogeny of southern hemisphere stoneflies: Testing for Gondwanan origins. *Molecular Phylogenetics and Evolution*, 96, 150–160. <https://doi.org/10.1016/j.ympev.2015.10.028>.
- Miller, M. A., Pfeiffer, W., & Schwartz, T. (2010). Creating the CIPRES Science Gateway for inference of large phylogenetic trees. *Proceedings of the Gateway Computing Environments Workshop (GCE)*, 1–8, 1–8. <https://doi.org/10.1109/GCE.2010.5676129>.
- Misof, B., Liu, S., Meusemann, K., Peters, R. S., Donath, A., Mayer, C., Frandsen, P. B., Ware, J., Flouri, T., Beutel, R. G., Niehuis, O., Petersen, M., Izquierdo-Carrasco, F., Wappler, T., Rust, J., Aberer, A. J., Aspöck, U., Aspöck, H., Bartel, D., ... Zhou, X. (2014). Phylogenomics resolves the timing and pattern of insect evolution. *Science*, 346, 763–767. <https://doi.org/10.1126/science.1257570>.
- Monjardim, M., Paresque, R., & Salles, F. F. (2020). Phylogeny and classification of Leptophlebiidae (Ephemeroptera) with an emphasis on Neotropical fauna. *Systematic Entomology*, 45, 415–429. <https://doi.org/10.1111/syen.12402>.
- Mueller, C. O., & Jokat, W. (2019). The initial Gondwana break-up: A synthesis based on new potential field data of the Africa-Antarctica Corridor. *Tectonophysics*, 750, 301–328. <https://doi.org/10.1016/j.tecto.2018.11.008>.
- O'Donnell, B. C., & Jockusch, E. L. (2008). Phylogenetic relationships of leptophlebiid mayflies as inferred by histone H3 and 28S ribosomal DNA. *Systematic Entomology*, 33, 651–667. <https://doi.org/10.1111/j.1365-3113.2008.00434.x>.
- Ogden, T. H., Breinholt, J. W., Bybee, S. M., Miller, D., Sartori, M., Shiozawa, D., & Whiting, M. (2019). Mayfly phylogenomics: Initial evaluation of anchored hybrid enrichment data for the order Ephemeroptera. *Zoosymposia*, 16, 167–181. <https://doi.org/10.11646/zoosymposia.16.1.14>.
- Parras, A., & Griffin, M. (2013). Late Cretaceous (Campanian/Maastrichtian) freshwater to restricted marine mollusc fauna from the Loncoche Formation, Neuquén Basin, west-central Argentina. *Cretaceous Research*, 40, 190–206. <https://doi.org/10.1016/j.cretres.2012.07.002>.
- Pescador, M. L., & Peters, W. L. (1980). Phylogenetic relationships and zoogeography of cool-adapted Leptophlebiidae (Ephemeroptera) in southern South America. In J. F. Flannagan & K. E. Marshall (Eds.), *Advances in ephemeroptera biology* (pp. 43–56). Springer.
- Pescador, M. L., & Peters, W. L. (1990). Biosystematics of the genus *Massartella* Lestage (Ephemeroptera: Leptophlebiidae: Atalophlebiinae) from South America. *Aquatic Insects*, 12, 145–160. <https://doi.org/10.1080/01650429009361399>.
- Pike, H. (2019). It's time to talk about ditching statistical significance. *Nature*, 567, 283. <https://doi.org/10.1038/d41586-019-00874-8>.
- R Core Team (2020). *R: A language and environment for statistical computing*. R Foundation for Statistical Computing. <https://www.R-project.org/>.
- Rambaut, A., Drummond, A. J., Xie, D., Baele, G., & Suchard, M. A. (2018). Posterior summarization in Bayesian phylogenetics using Tracer 1.7. *Systematic Biology*, 67, 901. <https://doi.org/10.1093/sysbio/syy032>.
- Ree, R. H., Moore, B. R., Webb, C. O., & Donoghue, M. J. (2005). A likelihood framework for inferring the evolution of geographic range on phylogenetic trees. *Evolution*, 59, 2299–2311.
- Ree, R. H., & Sanmartín, I. (2018). Conceptual and statistical problems with the DEC+J model of founder-event speciation and its comparison with DEC via model selection. *Journal of Biogeography*, 45, 741–749. <https://doi.org/10.1111/jbi.13173>.
- Ree, R. H., & Smith, S. A. (2008). Maximum likelihood inference of geographic range evolution by dispersal, local extinction, and cladogenesis. *Systematic Biology*, 57, 4–14. <https://doi.org/10.1080/10635150701883881>.
- Reguero, M. A., Gelfo, J. N., López, G. M., Bond, M., Abello, A., Santillana, S. N., & Marensi, S. A. (2014). Final Gondwana breakup: The Paleogene South American native ungulates and the demise of the South America-Antarctica land connection. *Global and Planetary Change*, 123, 400–413. <https://doi.org/10.1016/j.gloplacha.2014.07.016>.
- Ronquist, F., & Sanmartín, I. (2011). Phylogenetic methods in biogeography. *Annual Review of Ecology, Evolution, and Systematics*, 42, <https://doi.org/10.1146/annurev-ecolsys-102209-144710>.
- Ronquist, F., Teslenko, M., van der Mark, P., Ayres, D. L., Darling, A., Höhna, S., Larget, B., Liu, L., Suchard, M. A., & Huelsenbeck, J. P. (2012). MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology*, 61, 539–542. <https://doi.org/10.1093/sysbio/sys029>.
- Sanmartín, I., & Ronquist, F. (2004). Southern hemisphere biogeography inferred by event-based models: Plant versus animal patterns. *Systematic Biology*, 53(2), 216–243.
- Sartori, M., & Brittain, J. E. (2015). Order Ephemeroptera. In J. H. Thorp, & D. C. Roger (Eds.), *Thorp and Covich's freshwater invertebrates Volume 1. Ecology and general biology* (pp. 873–891). Academic Press.
- Savage, H. M. (1987). Biogeographic classification of the Neotropical Leptophlebiidae (Ephemeroptera) based upon geological centers of ancestral origin and ecology. *Studies on Neotropical Fauna and Environment*, 22, 199–222. <https://doi.org/10.1080/01650528709360734>.
- Scher, H. D., Whittaker, J. M., Williams, S. E., Latimer, J. C., Kordesch, W. E., & Delaney, M. L. (2015). Onset of Antarctic Circumpolar Current 30 million years ago as Tasmanian Gateway aligned with westerlies. *Nature*, 523, 580–583. <https://doi.org/10.1038/nature14598>.
- Scotese, C. R., & Dreher, C. (2012). GlobalGeology. <http://www.globalgeology.com>.
- Seton, M., Müller, R. D., Zahirovic, S., Gaina, C., Torsvik, T., Shephard, G., Talsma, A., Gurnis, M., Turner, M., Maus, S., & Chandler, M. (2012). Global continental and ocean basin reconstructions since 200 Ma. *Earth-Science Reviews*, 113, 212–270. <https://doi.org/10.1016/j.earscirev.2012.03.002>.
- Sroka, P., Staniczek, A. H., & Bechly, G. (2015). Revision of the giant pterygote insect *Bojophlebia prokopi* Kukalová-Peck, 1985 (Hydropalaeoptera: Bojophlebiidae) from the Carboniferous of the Czech Republic, with the first cladistic analysis of fossil palaeopteran insects. *Journal of Systematic Palaeontology*, 13, 963–982. <https://doi.org/10.1080/14772019.2014.987958>.
- Stadler, T. (2009). On incomplete sampling under birth-death models and connections to the sampling-based coalescent. *Journal of Theoretical Biology*, 261, 58–66. <https://doi.org/10.1016/j.jtbi.2009.07.018>.
- Storari, A., Rodrigues, T., Saraiva, A. A. F., & Salles, F. F. (2020). Unmasking a gap: A new oligoneuriid fossil (Ephemeroptera: Insecta) from the Crato Formation (upper Aptian), Araripe Basin, NE Brazil, with

- comments on *Colocrus* McCafferty. *PLoS ONE*, 15, e024036. <https://doi.org/10.1371/journal.pone.0240365>.
- Thompson, J. O., Moulin, M., Aslanian, D., De Clarens, P., & Guillocheau, F. (2019). New starting point for the Indian Ocean: Second phase of breakup for Gondwana. *Earth-Science Reviews*, 191, 26–56. <https://doi.org/10.1016/j.earscirev.2019.01.018>.
- Toussaint, E. F., Bloom, D., & Short, A. E. (2017). Cretaceous West Gondwana vicariance shaped giant water scavenger beetle biogeography. *Journal of Biogeography*, 44, 1952–1965. <https://doi.org/10.1111/jbi.12977>.
- Toussaint, E. F., Hendrich, L., Hájek, J., Michat, M. C., Panjaitan, R., Short, A. E., & Balke, M. (2017). Evolution of Pacific Rim diving beetles sheds light on Amphi-Pacific biogeography. *Ecography*, 40, 500–510. <https://doi.org/10.1111/ecog.02195>.
- Varela, A. N., Raigemborn, M. S., Richiano, S., White, T., Poiré, D. G., & Lizzoli, S. (2018). Late Cretaceous paleosols as paleoclimate proxies of high-latitude Southern Hemisphere: Mata Amarilla Formation, Patagonia, Argentina. *Sedimentary Geology*, 363, 83–95. <https://doi.org/10.1016/j.sedgeo.2017.11.001>.
- Wasserstein, R. L., Schirm, A. L., & Lazar, N. A. (2019). Moving to a world beyond “ $p < 0.05$ ”. *The American Statistician*, 73, 1–19. <https://doi.org/10.1080/00031305.2019.1583913>.
- White, L. T., Gibson, G. M., & Lister, G. S. (2013). A reassessment of paleogeographic reconstructions of eastern Gondwana: Bringing geology back into the equation. *Gondwana Research*, 24, 984–998. <https://doi.org/10.1016/j.gr.2013.06.009>.
- Xia, X. (2018). DAMBE7: New and improved tools for data analysis in molecular biology and evolution. *Molecular Biology and Evolution*, 35, 1550–1552. <https://doi.org/10.1093/molbev/msy073>.
- Xia, X., Xie, Z., Salemi, M., Chen, L., & Wang, Y. (2003). An index of substitution saturation and its application. *Molecular Phylogenetics and Evolution*, 26, 1–7. [https://doi.org/10.1016/S1055-7903\(02\)00326-3](https://doi.org/10.1016/S1055-7903(02)00326-3).

- Yu, Y., Harris, A. J., Blair, C., & He, X. J. (2015). RASP (Reconstruct Ancestral State in Phylogenies): A tool for historical biogeography. *Molecular Phylogenetics and Evolution*, 87, 46–49. <https://doi.org/10.1016/j.ympev.2015.03.008>.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

Figure S1. Phylogenetic relationships of Atalophlebiinae inferred by Bayesian analysis on a concatenated molecular dataset (28S+COI)

Table S1. List of Atalophlebiinae and outgroups used in this research, including GenBank accession numbers and voucher numbers

Table S2. Details of the primers used in this research

Table S3. Best substitution models inferred by jModelTest2 using Corrected Akaike Information Criterion (AICc)

Table S4. Probability of dispersal according to the availability of connections among areas across four time slices. South America (A), Madagascar (B), Australia (C), and New Zealand (D)

Alignment S1. Alignment of sequences included in this research

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